



Original Article

## Density, recruitment and growth performance of Asian green mussel (*Perna viridis*) in Marudu Bay, Northeast Malaysian Borneo, three years after a massive mortality event

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### Abstract

Density, recruitment and growth performance of Asian green mussel (*Perna viridis*) in a particular coastal marine environment can be affected by many factors, including environmental change, pollution, disease outbreak and massive mortality event. The present study was conducted to determine the density, recruitment and growth performance of farmed Asian green mussel in Marudu Bay, three years after a mass mortality event. The study was carried out for 12 months between April 2013 and March 2014. The length frequency data of 1,308 individuals of green mussel were analyzed using the latest version of the FAO-ICLARM Fish Stock Assessment Tools (FiSAT II). The result showed that the green mussel recruitment in Marudu Bay occurs throughout the year with two major peaks i.e. February and July which coincided with the monsoon seasons. The asymptotic length ( $L_{\infty}$ ), growth coefficient (K) and growth performance index ( $\phi'$ ) of the farmed Asian green mussel in Marudu Bay are relatively high at 113.4 mm, 1.7 year<sup>-1</sup> and 4.34, respectively. However, despite good culture location, the settlement density of green mussel in the bay was low. We suspected that the low settlement density could be influenced by the ecological effects due to the long term suspension of the culture substrates and the physiochemical properties of the water in Marudu Bay. Nevertheless, chlorophyll-*a* measurement alone was not able to justify if food scarcity has resulted in high mortality of the farmed Asian green mussel in Marudu Bay.

**Keywords:** mussels, *Perna viridis*, density, recruitment, growth performance

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### 1. Introduction

Asian green mussel (*Perna viridis*) is an important seafood resource and it is widely cultivated for commercial purposes especially in the Southeast Asian region. This species is extensively cultured due to its high productivity, high tolerance to a wide range of environmental conditions, and it requires less farm management (Rajagopal *et al.*, 2006; Al-Barwani *et al.*, 2007; McFarland *et al.*, 2013). *Perna viridis* is currently being recognized as a cheap protein sources, containing high nutritional values and it is popular for its

delicious taste (Rajagopal *et al.*, 1998; Yap, 2012).

The production of green mussel in Malaysia reached the highest peak in 2010 with total production of 10,529.06 MT, but declined continuously until 2013 (1,070.88 MT) by 89.9% reduction compared to that in 2010 (DOF, 2010; DOF, 2013). One of the major causes of the decline was the occurrence of massive mortality event in many green mussel farms across the country. One of these farms was located in Marudu Bay, northeast Malaysian Borneo. The green mussel aquaculture was introduced in the bay in the late 1990s and became a commercially important activity in early 2000. Unfortunately, in late 2009 to 2012 the green mussel farm in Marudu Bay was seriously affected by massive mortality. The mortality event wiped out almost all the juveniles and adults mussel population, leaving only small quantity of survived

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mussels on culture ropes. Since then, the production of green mussel in the bay had drastically gone down, deserted and caused huge economic loss to farmers.

Previous studies described several factors which can cause mussel mortality including physiochemical, hydrodynamic, food, predation, and diseases outbreaks (Gulshad, 2003; Schiel, 2004; Peperzak and Poelman, 2008; Yap, 2012; Heinonen, 2014). Among these causes of mortality events, the environmental parameters and food availability are the most reported causes of massive mortality of farmed bivalve. For example, sudden increase in water temperature is often lead to mortality of green mussel and other bivalve species under experimental conditions (Hiebenthal *et al.*, 2012; Sreedevi *et al.*, 2014; Solomieu *et al.*, 2015). Furthermore, Alforo (2006) found the mortality of *Perna canaliculus* in northern New Zealand was due to limited food supply.

The of physiochemical parameters of water, water nutrients and chlorophyll are essential to establish the relationship between growth and abundance of green mussel in Marudu Bay and the environmental factors. Besides, information pertaining to recovery rates such as recruitment, growth and mortality after a mass mortality event is also essential for farm management to understand the extent of the potential risk and thus possible mitigation for future restocking. In this study, we investigated the density, recruitment pattern and growth performance of green mussel cultured using the hanging rope culture method in Marudu Bay, three years after the mass mortality event.

## 2. Materials and Methods

### 2.1 Study area

This study was conducted in a green mussel farm in Marudu Bay, (6°38'22" N, 116°53'17" E), Sabah, Malaysia; Figure 1). Marudu Bay is influenced by two monsoons, the

Northeast Monsoon that occurs between November and April and the Southwest Monsoon that takes place between May and September. Heavy rainfall generally occurs during the Northeast Monsoon.

### 2.2 Field experiment and sampling

The experiment was carried out for 12 months from April 2013 to March 2014. The Asian green mussel was cultured using the hanging rope culture method. Nylon ropes (length x diameter: 1 m x 2 cm) wrapped with a fine filament of fish netting were used as substrate to facilitate the settlement of the mussel spats. A total of 250 ropes were hung on the raft and suspended at 0.5 m depth below the water surface with 30 cm rope spacing. Samplings were conducted once a month. Ropes with nine replicates were collected randomly on a monthly interval to estimate the density of the mussel. All analyses on the mussel samples were conducted within 48 hours of collection.

### 2.3 Length and weight measurement

In the laboratory, the ropes were washed with running seawater and the mussels were individually removed. Morphometric measurements including length, thickness and weight were measured according to the methods described by Vakily *et al.* (1988) using a caliper at 0.1 mm accuracy. The total weight was weighed by using analytical balance (Sartorius) of 0.001 gram accuracy. In total 1,308 mussel samples were then grouped into shell length classes of 5 mm interval following Al-Barwani *et al.* (2007).

### 2.4 Water physiochemical parameters

Physiochemical parameters of water such as dissolve oxygen (DO), pH, salinity and temperature was measured

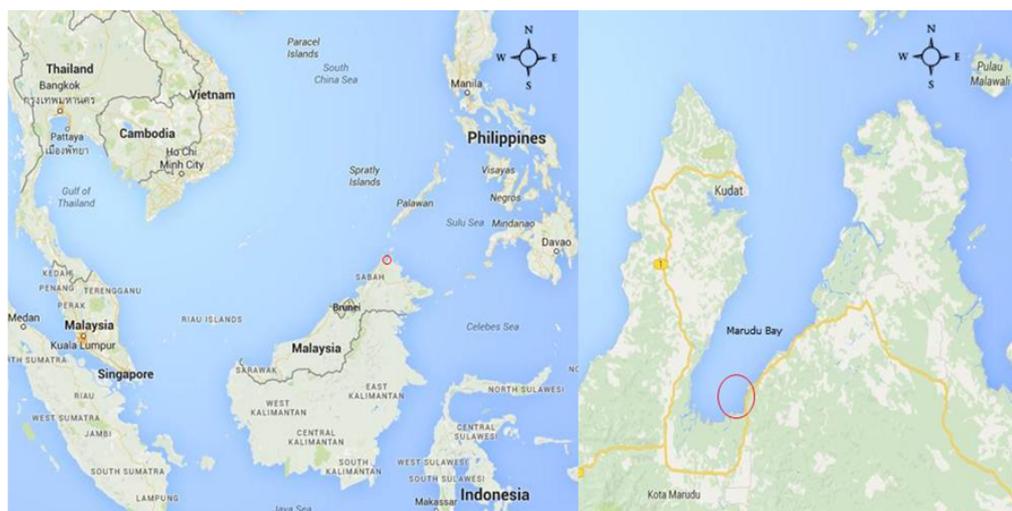


Figure 1. Map shows the Marudu Bay (red circle) on the northeastern of Malaysian Borneo (left map). Approximate location of the sampling site (red circle) within the Marudu Bay (right map). Source: Google Maps.

once every month by using a HI 9829 Multi-parameters water quality checker (Hanna Instrument). Current speed was measured by using the Direct Reading Electromagnetic Current Meter (AEM-213D).

**2.5 Water nutrients and chlorophyll- $\alpha$  analysis**

In total of 500 ml of water samples were filtered through membrane filter (0.45 $\mu$ m) with the help of a vacuum pump. The filtered membranes were subjected to chlorophyll- $\alpha$  analysis following the method of Strickland and Parsons (1972). Chlorophyll- $\alpha$  was then extracted with 10 ml of 90% acetone. The absorbance of the extracts was determined at 664, 647, and 630 nm. The chlorophyll- $\alpha$  concentration was calculated by following equation (Talling and Driver, 1963):

Chlorophyll- $\alpha$  ( $\mu$ g/L) =

$$(11.85 A_{664} - 1.54 A_{647} - 0.08 A_{630}) \times \frac{V}{S} \times 1000$$

$A_{664}$  = Absorbance at 664 nm

$A_{647}$  = Absorbance at 647 nm

$A_{630}$  = Absorbance at 630 nm

V = Volume of acetone used (mL)

S = Volume of sampled filter (mL)

In the other hand, 25 ml each of the filtered seawater was analyzed for water nutrients (ammonia, nitrate, nitrite and phosphate) following the method of Parsons *et al.* (1984). The concentration of each water nutrient was determined spectrophotometric at 640 nm (ammonia), 543 nm (nitrate and nitrite), and 882 nm (phosphate).

**2.6 Statistical analysis**

Growth and recruitment of farmed Asian green mussel at different months were analyzed by ONE-way ANOVA followed by Turkey multiple comparison test (Turkey HSD). Data were also subjected to correlation and bivariate tests to find significant relationship between the mussel density and water parameters. Tests were judged to be significant at  $p < 0.05$  level. Prior to analyses, all the variables were tested for normality and homogeneity of variances using One-way ANOVA. All the parametric tests were performed by using SPSS Windows Statistical Package (version 18).

Von Bertalanffy growth function (VBGF) and recruitment pattern of green mussel was estimated based on frequency distribution of each length class sizes every month for one year period. The estimation were performed in ELEFAN-1 (Pauly and David, 1981) using the FiSAT software as explained in detail by Gayanilo *et al.* (1996). Asymptotic length ( $L_{\infty}$ ) and growth coefficient ( $K$ ) of the  $K$ -scan routine was conducted to assess a reliable estimate of the  $K$  value. The parameters  $L_{\infty}$  and  $K$  were then used to calculate the growth performance index ( $\phi'$ ) of the farmed Asian green mussel using the equation  $\phi' = 2 \text{Log}_{10} L_{\infty} + \text{Log}_{10} K$  (Pauly and Munro, 1984). Normal distribution of the recruitment pattern

was determined by using the NORMSEP, FiSAT (Pauly and Caddy, 1985). On the other hand, the density of the mussel in 1 m<sup>2</sup> surface area of the substrate (rope) was estimated using the following formula.

Density (per m<sup>2</sup>) =

$$\frac{\text{Number of enumerated individuals (N)}}{\text{Surface area of substrate (rope) } (2\pi r^2 + 2\pi r(h))}$$

where N = number of green mussel in 1 rope (total individual found /9 ropes taken every month), h = 1 m (height of rope), and r = 0.01m (radius of rope).

**3. Results**

**3.1 Mussel density**

The monthly density of the Asian green mussels attached to substrate (ropes) is illustrated in Figure 2. Highest number of the mussels (about 500 ind m<sup>-2</sup>) was recorded in December 2013. This value was significantly higher ( $p < 0.05$ ) than the density of the mussels recorded in any other months (150 to 250 ind m<sup>-2</sup>).

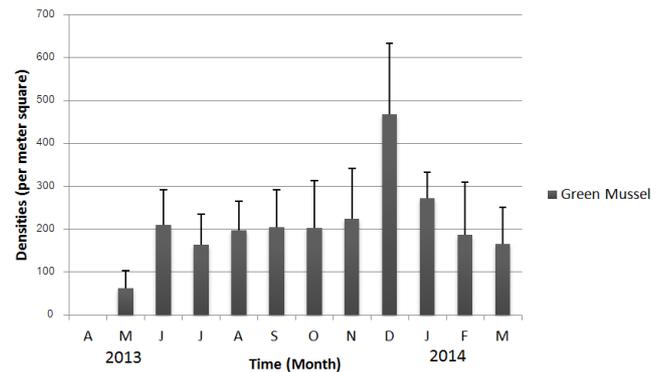


Figure 2. Density of farmed Asian green mussel attached on the experimental substrate (ropes) in Marudu Bay recorded from April 2013 to March 2014. Standard error (SE) is indicated by the bar line on top of each bar.

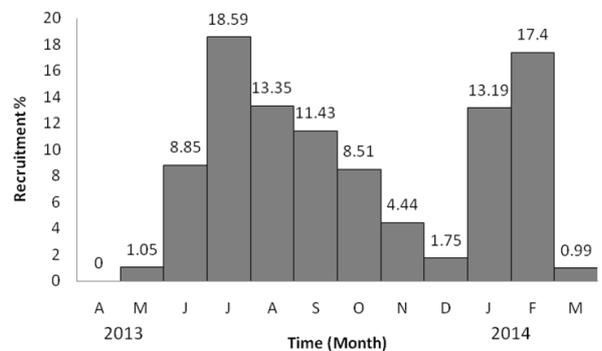


Figure 3. Recruitment percentage of farmed Asian green mussel recorded in Marudu Bay between April 2013 and March 2014. The recruitment was estimated based on the time projection of length frequencies.

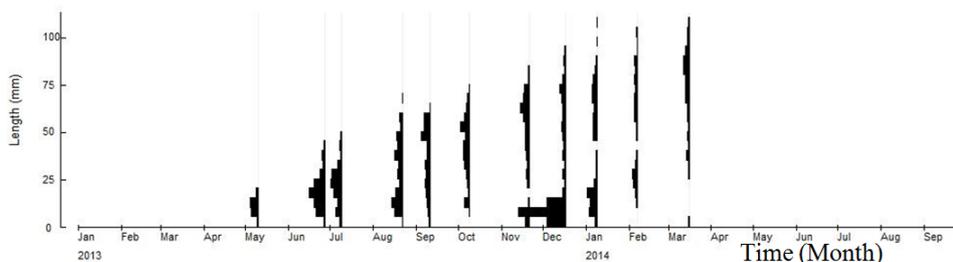


Figure 4. The length frequency data of farmed Asian green mussel in Marudu Bay. Note that the class sizes spread widely and oscillate every month.

### 3.2 Mussel recruitment

Recruitment of Asian green mussel occurred throughout the year with two seasonal peaks (Figure 3). The peaks recruitment of 18% and 17% were recorded in July 2013 and February 2014, respectively. The cohorts arising from the July recruitment were found to reach a marketable size (50-60 mm) in 5 to 6 months (Figure 4).

### 3.3 Growth performance

The asymptotic length ( $L_{\infty}$ ) of the farmed Asian green mussel was estimated at 113.4 mm and the growth coefficient (K) was at 1.70 year<sup>-1</sup> (Figure 5). The calculated growth performance index ( $\phi'$ ) was 4.34. The maximum length observed in 12 months monitoring was 110 mm, and the predicted extreme length of the mussel was 129.03mm. The maximum length at 95% confidence range was 102.91-155.15 mm (Figure 6). The monthly average length and weight of the mussels was relatively increased in tandem with the study period. In December 2013, the average length and weight dropped more than 50% compared to November 2013. The highest average length and weight recorded in the culture area was in March 2014 with 82% (74.6mm) increase in length and 99% (30.6g) increase in weight compared to that in May 2013.

### 3.4 Relationship between mussel density and water parameters

The water parameters recorded throughout the one-year sampling period are shown in Figure 7. The highest surface water temperature was recorded in April 2013, while the lowest water temperature recorded in October 2013. The salinity varied from 25 ppt to 31 ppt. Meanwhile, water pH recorded during the sampling period was within the normal range (7.6-8.2). Dissolved oxygen fluctuated from 3.5 mg/L (May 2013) to 5.9 mg/L (January 2014). Water velocity (current speed) exhibited variation from 5.85 cm/s to 19.96 cm/s. Water nutrients levels were relatively high between April 2013, and December 2014 to March 2014. However, low averages of nitrate and nitrite concentrations (below 1  $\mu\text{g/L}$ ) were recorded throughout the sampling period. Chlorophyll- $\alpha$  concentration was recorded slightly higher in June and October but low between December and April. The bivariate

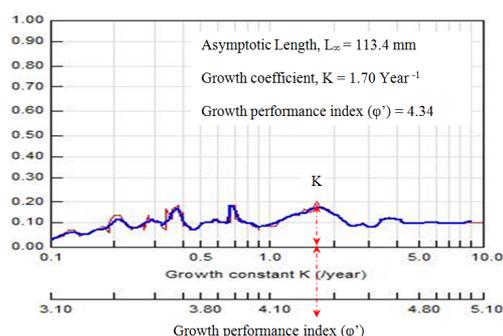


Figure 5. Von Bertalanffy growth function of farmed Asian green mussel in Marudu Bay estimated between April 2013 and March 2014.

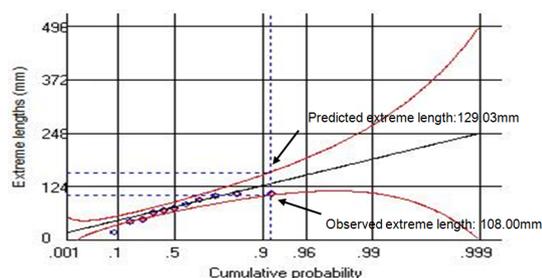


Figure 6. Maximum length of farmed Asian green mussel in Marudu Bay predicted from extreme values, with range at 95% confidence interval: 102.91mm - 155.15mm.

test illustrated that there was a significantly negative relationship between the mussel density with water temperature, velocity and phosphate concentrations ( $p < 0.01$ ) as shown in Table 1. However, the mussel density was found to have a significant positive correlation with dissolved oxygen.

## 4. Discussion

The highest density of *Perna viridis* was only observed in certain months (December 2013 and January 2014). Similar finding was also reported in India and Thailand during the first year of substrate deployment (Chaitanawisuti and Menasveta, 1987; Rajagopal *et al.*, 1998). However, the density of green mussel in study area was extremely lower than reported in Hong Kong, Thailand, New Zealand and

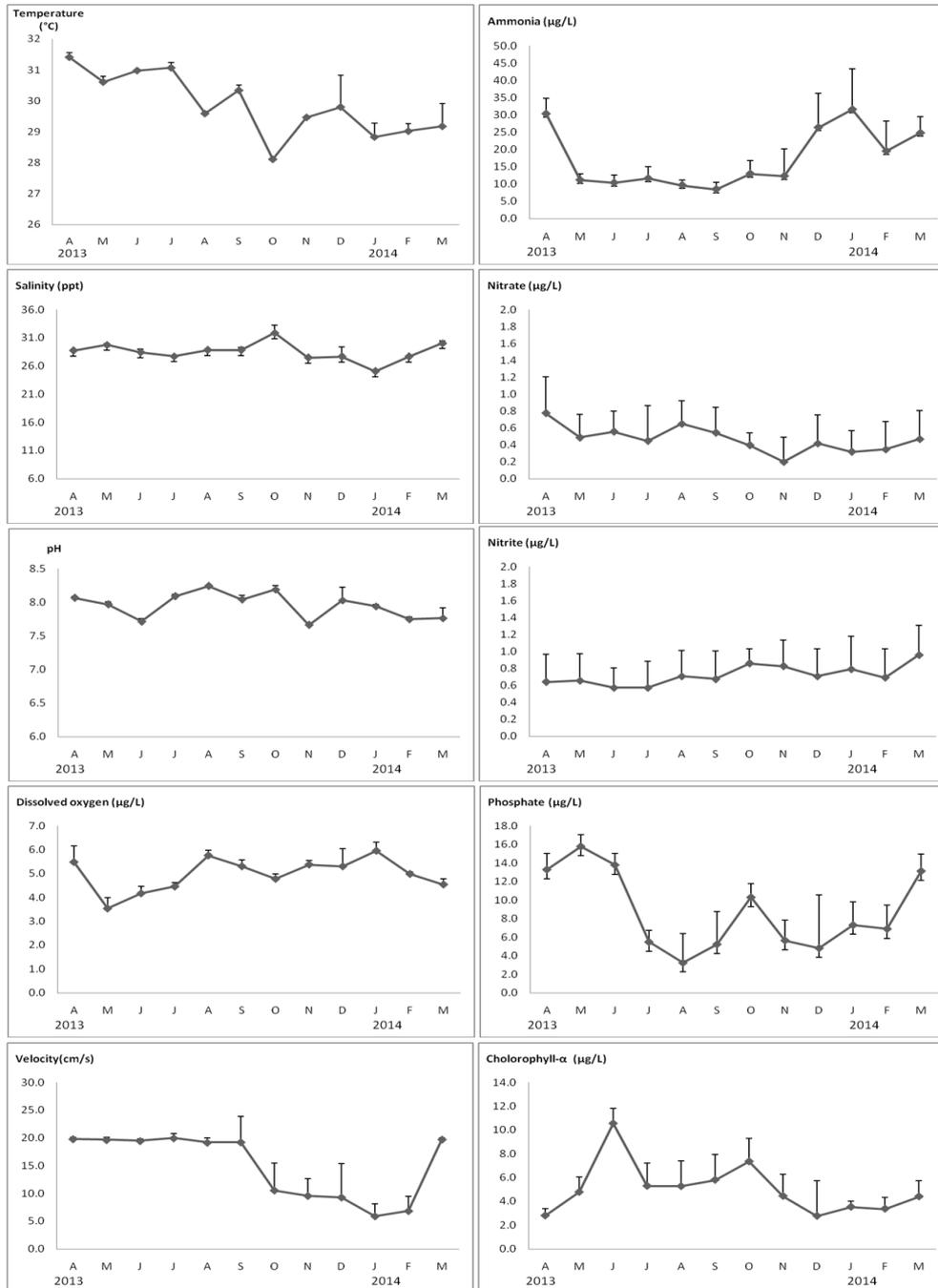


Figure 7. Water parameters in Marudu Bay recorded from April 2013 to March 2014.

India mussel farms with settlement density more than 1,000 ind m<sup>-2</sup> (Lee, 1985; Chaitanawisuti and Menasveta, 1987; Rajagopal *et al.*, 1998; Alfaro, 2006). Despite the poor density, the recruitment was found to occur all-year round, similar to that in other tropical Asian countries (Al-Barwani *et al.*, 2007; Khan *et al.*, 2010; Laxmilatha, 2013). The major recruitment of the green mussel in Marudu Bay occurred twice a year; first in July (Southwest Monsoon) and second in February (Northeast Monsoon) which coincided with the monsoon seasons in Malaysia. Although recruitment of the mussel

occurs throughout the year, its peaks of recruitment vary according to places (Khan *et al.*, 2010). Such phenomenon may be influenced by the complicated interactions between biological, chemical and physical factors (Broitman *et al.*, 2005; Smith *et al.*, 2009).

In the present study, the asymptotic length ( $L_{\infty}$  = 113.4mm) of the farmed green mussel in Marudu Bay was recorded higher than those reported in other places in Malaysia including Malacca (102.38) and Penang (89.4mm) (Al-Barwani *et al.*, 2007). This value is resembled to those

Table 1. Pearson correlation coefficient (r) between farmed Asian green mussel density and water parameters

	Density	Temperature	pH	Salinity	Dissolved oxygen	Velocity	Ammonia	Nitrate	Nitrite	Phosphate	Chlorophyll- $\alpha$
Density	1	<b>-0.455**</b>	-0.179	-0.285	<b>0.367*</b>	<b>-0.507**</b>	-0.082	-0.287	0.158	<b>-0.336*</b>	-0.060
Temperature	<b>-0.455**</b>	1	0.106	0.013	-0.362*	0.759**	-0.007	0.410*	-0.149	0.237	-0.050
pH	-0.179	0.106	1	0.306	0.075	0.329	-0.173	0.351*	-0.069	-0.281	0.330*
Salinity	-0.285	0.013	0.306	1	-0.682**	0.468**	-0.267	0.091	0.110	0.333*	0.467**
Dissolved oxygen	<b>0.367*</b>	-0.362*	0.075	-0.682**	1	-0.471**	0.316	0.041	-0.077	-0.588**	-0.335*
velocity	<b>-0.507**</b>	0.759**	0.329	0.468**	-0.471**	1	-0.227	0.446**	-0.145	0.210	0.320
Ammonia	-0.082	-0.007	-0.173	-0.267	0.316	-0.227	1	0.307	0.345*	0.225	-0.627**
Nitrate	-0.287	0.410*	0.351*	0.091	0.041	0.446**	0.307	1	0.149	0.185	0.012
Nitrite	0.158	-0.149	-0.069	0.110	-0.077	-0.145	0.345*	0.149	1	0.304	-0.148
Phosphate	<b>-0.336*</b>	0.237	-0.281	0.333*	-0.588**	0.210	0.225	0.185	0.304	1	-0.052
Chlorophyll- $\alpha$	-0.060	-0.050	0.330*	0.467**	-0.335*	0.320	-0.627**	0.012	-0.148	-0.052	1

\*\*correlation is significant at the 0.01 level (2-tailed); \*correlation is significant at the 0.05 level (2-tailed).

reported in other Asian countries particularly in Hong Kong and Thailand at 101.9 mm and 112 mm, respectively (Lee, 1985; Tuaycharden *et al.*, 1988). The  $K$  value of the mussel in Marudu Bay was higher compared to those in Malacca (Albarwani *et al.*, 2007), Bangladesh (Khan *et al.*, 2010), Hong Kong (Lee, 1985), India (Narasimham, 1981) and Thailand (Tuaycharden *et al.*, 1988). The high  $K$  value and the excellent asymptotic length ( $L_{\infty}$ ) of the mussel in Marudu Bay may be explained by the rapid growth of mussels in small clump with less density ( $\sim 9$  ind), than the mussels in larger clumps with high density ( $< 20$  ind) (Seed and Suchanek, 1992).

The physiochemical properties of seawater in Marudu Bay can be characterized as suitable site for mussel farming because they ranged within the suggested good conditions for mussels (Shamsudin, 1992; Kingzett and Salmon, 2002; Tan and Ransangan, 2014). Massive mortality event in late 2009 occurred approximately after 10 years since the introduction of green mussel aquaculture in Marudu Bay has caused significant reduction in commercial production. After three years of the mortality event, density of the green mussel on suspended ropes (substrate) is still low. Such situation might be best explained by the ecological effects of the long term suspension of the ropes used for mussel farming (Keeley *et al.*, 2009; Yap, 2012) and adaptation of the introduced green mussels to the ambient environment in the bay (Riisgard *et al.*, 2013; Zbawicka *et al.*, 2014). Ecological effects create novel habitats for micro and macro fouling organism such as algae (focus) and barnacles to outcompete for space and food (Bendell-Young, 2006; Yap, 2012). Colonization of other biofouling and mussel epibionts on suspension ropes as well as on mussel shell influences the mussel growth and may cause poor mussel settlement (Garner and Litvaitis, 2013a). Weak settlement increases the risk of dislodgement and detachment from the substratum and may lead to high mortality (Harger and Landenberger, 1971; Yap, 2012).

Increase in temperature and water velocity has been reported to cause disturbances to mussel settlement on rope

substrates by means of byssogenesis (Tamarin *et al.*, 1974). Strength of byssal threads is important for green mussel to remain anchored on the suspension substrate (Garner and Litvaitis, 2013b). *Mytilus edulis* (Carrington, 2002) and *Mytilus galloprovincialis* (Zardi *et al.*, 2007) were found to produce less byssal threads at high temperature and during reproductive period. Such occurrence is influenced by the high decaying rate and least energy provided for byssogenesis as mussels are allocating more energy for gamete production (Rajagopal *et al.*, 1998). In addition, at higher temperature ( $> 29^{\circ}\text{C}$ ) time taken by mussel larvae to settle on substrate becomes longer than at lower temperature ( $26^{\circ}\text{C}$ ) (Siddall, 1979; Monaj and Appukutan, 2003). Delay in settlement elevates the changes of mussel larvae and even adults to predation and mortality (Harger and Landenberger, 1971; Smale and Buchan, 1981).

Strong mechanical forces as the result of high water flow also suppressed the byssal threads production by weakening the mussel foot ability to produce strong byssal threads (Tamarin *et al.*, 1974; Carrington, 2002; Moeser *et al.*, 2006; Carrington *et al.*, 2008; Garner and Litvaitis, 2013a). This causes the mussels to experience difficulty in settling down on substrate. This was evident in the current study where the density of green mussels was higher ( $400$  ind  $\text{m}^{-2}$ ) in December 2013 compared to that in March 2014 ( $> 200$  ind  $\text{m}^{-2}$ ) with different water current recorded at  $9.7$  cm/s and  $19.8$  cm/s, respectively. According to Moeser *et al.* (2006), ambient water velocity for stronger byssal thread attachment of blue mussel was at  $11$  cm/s but reduced at above  $15$  cm/s. Combination of high water temperature and water velocities ( $r = 0.759$ ) in Marudu Bay could have influenced the mussel attachment resulting lower settlement density.

The low dissolved oxygen in Marudu Bay ( $r = 0.588$ ) could have been influenced by the high phosphorus content in the water column (McCormik and Laing, 2003). The high loading of phosphorus in the bay is originated from the intense anthropogenic activities which are recently taking place along the major rivers (Aris *et al.*, 2014) and the runoffs

from palm oil plantations (Zakaria and Rajpar, 2015) surrounding the bay. In this study, reduced dissolved oxygen and high phosphorus in water column correlated with reduced mussel settlement density. Low mussel population at high phosphorus content and low dissolved oxygen has also been reported by Sarnelle *et al.* (2012) and Clarke and McMahon (1996). There, mussel attachment strength on substrate surface may weaken under low dissolved oxygen (Clarke and McMahon, 1996), whereas high phosphate level reduces the mussel movement, ventilation and siphons closure. Such conditions can cause physiological stress to mussels (Jenner *et al.*, 1992; Reynolds and Guillaume, 1998).

Researchers have shown that there is a strong relationship between food availability and green mussel density (Rajagopal *et al.*, 1998; Alfaro, 2006). However, the present study did not find any correlation between monthly chlorophyll- $\alpha$  measurement and the settlement density of green mussel in the bay. This shows that the chlorophyll- $\alpha$  measurement alone is not able to justify the food sufficiency for bivalve due to their selective feeding behavior (Ren *et al.*, 2000; Rouillon and Navarro, 2003).

## 5. Conclusions

The high asymptotic length of the green mussel in Marudu Bay, although is influenced by the low density, it also indicates that the bay is good for mussel farming. Recruitment of green mussel in Marudu Bay occurs throughout the year with two seasonal peaks (July and February), one in the Southeast Monsoon and another one in Northeast Monsoon. The low density of the green mussel in the bay may have been influenced by the ecological effects due to long term suspension of the culture substrates. Increase in water temperature and water velocity may disturb or delay the attachment of green mussel to settle on substrates, hence resulting in low settlement density. The low dissolved oxygen and high amount of phosphorus in the bay may also affect the settlement density. Studies on the feeding behaviors and food preference of green mussel are necessary to determine if the high mortality of the green mussel in the bay is related to food scarcity.

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## References

- Al-Barwani, S.M., Arshad, A., Nurul Amin, S.M., Japar, S.B., Siraj, S.S., and Yap, C.K. 2007. Population Dynamics of the Green Mussel *Perna viridis* from the High Spat-fall Coastal Water of Malacca, Peninsular Malaysia. Fisheries Research. 84, 147–152.
- Alfaro, A.C. 2006. Population dynamics of the green-lipped mussel, *Perna canaliculus*, at various spatial and temporal scales in northern New Zealand. Journal of Experimental Marine Biology and Ecology. 334, 294-315.
- Aris, A.Z., Wan, Y.L., Praveena, S.M., Yusoff, M.K., Ramli, M. F., and Juahir, H. 2014. Water Quality Status of Selected Rivers in Kota Marudu, Sabah, Malaysia and its Suitability for Usage. Sains Malaysiana. 43(3), 377-388.
- Bendell-Young, I. 2006. contrasting the community structure and select geochemical characteristic of the three intertidal regions in relation to shellfish farming. Environmental Conservation. 33, 21-27.
- Broitman, B.R., Blanchette, C.A., and Gaines, S.D. 2005. Recruitment of intertidal invertebrates and oceanographic variability at Santa Cruz Island, California. Limnology and Oceanography. 50(5), 1473-1479.
- Carrington, E. 2002. Seasonal variation in the attachment strength of blue mussels: causes and consequences. Limnology and Oceanography. 47, 1723-1733.
- Carrington, E., Moeser, G.M., Thompson, S.B., Coutts, L.C., and Craig, C.A. 2008. Mussel attachment on rocky shores: the effect of ow on byssus production. Integrative and Comparative Biology. 48, 801–807.
- Chaitanawisuti, N. and Menasveta, P. 1987. Experimental Suspended Culture of Green Mussel, *Perna viridis* (Linn.), Using Spat Transplanted from a Distant Settlement Ground in Thailand. Aquaculture. 66, 97-107.
- Clarke, M. and McMahon, R.F. 1996. Effects of Hypoxia and Low-Frequency Agitation on Byssogenesis in the Freshwater Mussel *Dreissena polymorpha* (Pallas). Biological Bulletin. 191, 413-420.
- Department of Fisheries. 2010. Annual Fisheries Statistics 2008. Kuala Lumpur: Department of Fisheries, Malaysia.
- Department of Fisheries. 2013. Annual Fisheries Statistics 2010. Kuala Lumpur: Department of Fisheries, Malaysia.
- Garner, Y.L. and Litvaitis, M.K. 2013a. Effects of wave exposure, temperature and epibiont fouling on byssal thread production and growth in the blue mussel, *Mytilus edulis*, in the Gulf of Maine. Journal of Experimental Marine Biology and Ecology. 440, 52-56.
- Garner, Y.L. and Litvaitis, M.K. 2013b. Effects of injured conspecifics and predators on byssogenesis, attachment strength and movement in the blue mussel, *Mytilus edulis*. Journal of Experimental Marine Biology and Ecology. 448, 136-140.
- Gayanilo, F.C., Sparre, P., and Pauly, D. 1996. The FAO-ICLARM Stock Assessment Tools (FiSAT) users guide, FAO computerized information series, fisheries, FAO, Rome, Italy, pp. 126.
- Gulshad, M. 2003. Algal bloom and mass mortality of fishes and mussels along Kozhikode coast. Marine Fisheries Information Service, Technical and Extension Series, 175. pp. 7-8.

- Harger, J.R.E. and Landenberger, D.E. 1971. The Effect of Storms as a Density Dependent Mortality Factor on Populations of Sea Mussels. *Veliger*. 14(2), 195-201.
- Hiebenthal, C., Philipp, E.R., Eisenhauer, A., and Wahl, M. 2012. Interactive Effects of Temperature and Salinity on Shell Formation and General Condition in Baltic Sea *Mytilus edulis* and Arctic *Arctica islandica*. *Aquatic Biology*. 14, 289-298.
- Heinonen, K. 2014. Reports of Farmed Mussels *Mytilus* spp., *Perna* spp. Monterey Bay Aquarium's Seafood Watch, pp 30.
- Jenner, H.A., Van Aerssen, G.H.F.M., and Terwoert, J. 1992. Valve Movement Behaviour of the Mussel *Dreissena polymorpha* and the clam *Unio pictorum* for Use in an Early Warning System. *Limnologie Aktuell*. 4, 115-26.
- Keeley, N., Forrest, B., Hopkins, G., Gillespie, P., Clement, D., Webb, S., Knight, B.R., and Gardner, J. 2009. Sustainable aquaculture in New Zealand: Review of the ecological effects of farming shellfish and other non-fish species. Prepared for Ministry of Fisheries. Cawthron Report 1476. Cawthron Institute, Nelson, New Zealand.
- Khan, M.A., Assim, Z.B., and Ismail, N. 2010. Population Dynamics of the Green-Lipped Mussel, *Perna viridis* from the offshore Waters of Naf River Coast, Bangladesh. *Chiang Mai Journal of Science*. 37(2), 344-354.
- Kingzett, B. and Salmon, R. 2002. First Nation Shellfish Aquaculture Regional Business Strategy. Kingzett Professional Service Ltd., British Columbia, Canada.
- Laxmilatha, P. 2013. A Review of the Green Mussel *Perna viridis* Fishery of South West Coast of India. *International Journal of Marine Science*. 3(48), 408-416
- Lee, S.Y. 1985. The population dynamics of the green mussel, *Perna viridis* (L.) in Victoria Harbour, Hong Kong-dominance in a polluted environment. *Asian Marine Biology*. 2, 107-118.
- Manoj Nair, R. and Appukuttan, K.K. 2003. Effect of temperature on the development, growth, survival and settlement of green mussel *Perna viridis* (Linnaeus, 1758). *Aquaculture Research*. 34 (12), 1037-1045.
- McCormick, P.V. and Laing, J.E. 2003. Effects of increased phosphorus loading on dissolved oxygen in a subtropical wetland, the Florida Everglades. *Wetlands Ecology and Management*. 11, 199-216.
- McFarland, K., Donaghy, L., and Volety, A.K. 2013. Effect of acute salinity changes on hemolymph osmolality and clearance rate of the non-native mussel, *Perna viridis*, and the native oyster, *Crassostrea virginica*, in South-west Florida. *Aquatic Invasions*. 8(3), 299-310.
- Miller, B.A. and Emler, R.B. 1997. Influence of nearshore hydrodynamics on larval abundance and settlement of sea urchins *Strongylocentrotus franciscanus* and *S. purpuratus* in the Oregon upwelling zone. *Marine Ecology Progress Series*. 148, 83-94.
- Mooser, G.M., Leba, H., and Carrington, E. 2006. Seasonal influence of wave action on thread production in *Mytilus edulis*. *The Journal of Experimental Biology*. 209, 881-890.
- Narasimham, K.A. 1981. Dimensional relationships and growth of green mussel *Perna viridis* in Kakinada Bay. *Indian Journal of Fisheries*. 28, 240-248.
- Parsons, T. R., Marita, P., and M. Calli, M. 1984. A Manual of Chemical and Biological method for Seawater Analysis. Pergamon Press, U.K.
- Pauly, D. and David, N. 1981. ELEFAN-I BASIC program for the objective extraction of growth parameters from length-frequency data. *Meeresforschung*. 28(4), 205-211.
- Pauly, D. and Munro, J.L. 1984. Once more on the comparison of growth in fish and invertebrate. *International Center for Living Aquatic Resources Management Fishbyte*. 2(1), 21.
- Pauly, D. and Caddy, J.F. 1985. A modification of Bhattacharya's method for the analysis of mixtures of normal distributions. *FAO Fisheries Circular*, Vol. 781. FAO, Rome, Italy, pp 16.
- Peperzak, L. and Poelman, M. 2008. Mass mussel mortality in The Netherlands after a bloom of *Phaeocystis globosa* (prymnesiophyceae). *Journal of Sea Research*. 60, 220-222.
- Rajagopal, S., Venugopalan, V.P., Nair, K.V.K., Van der Velde, G., and Jenner, H.A. 1998. Settlement and growth of the green mussel *Perna viridis* (L.) in coastal waters: influence of water velocity. *Aquatic Ecology*. 32, 313-322.
- Rajagopal, S., Venugopalan, V.P., Van der Velde, G., and Jenner, H.A. 2006. Mussel colonization of a high low artificial benthic habitat: Byssogenesis holds the key. *Marine Environmental Research*. 62, 98-115.
- Ren, J.S., Ross, A.H., and Hayden, B.J. 2000. Comparison of Assimilation Efficiency on Diets of Nine Phytoplankton Species of the Greenshell Mussel *Perna canaliculus*. *Journal of Shellfish Research*. 25, 887-892.
- Reynolds, J. D. and Guillaume, H. P. 1998. Effects of phosphate on the reproductive symbiosis between bitterling and freshwater mussels: implications for conservation. *Journal of Applied Ecology*. 35, 575-581.
- Riisgard, H.U., Luskow, F., Pleissner, D., Lundgreen, K., and Lopez, M.A.P. 2013. Effect of salinity on filtration rates of mussels *Mytilus edulis* with special emphasis on dwarfed mussels from the low-saline Central Baltic Sea. *Helgoland Marine Research*. 67, 591-598.
- Rouillon, G. and Navarro, E. 2003. Differential Utilization of Species of Phytoplankton by the Mussel *Mytilus edulis*. *Acta Oecologica*. 24, 299-305.
- Sarnelle, O., White, J.D., Horst, G.P., and Hamilton, S.K. 2012. Phosphorus addition reverses the positive effect of zebra mussels (*Dreissena polymorpha*) on the toxic cyanobacterium, *Microcystis aeruginosa*. *Water Research*. 46, 3471-3478.

- Schiel, D.R. 2004. The structure and replenishment of rocky shore intertidal communities and biogeographic comparisons. *Journal of Experimental Marine Biology and Ecology*. 300, 309–342.
- Seed, R. and Suchanek, T.H. 1992. Population and community ecology of *Mytilus*. In: Gosling, E. (Ed.), *The Mussel Mytilus: Ecology, Genetics and Culture*. Elsevier, Amsterdam. 87-157.
- Shamsudin, L. 1992. *Akuakultur Pinggir Laut*. Dewan Bahasa dan Pustaka. Kementerian Pertanian Malaysia, Kuala Lumpur.
- Siddall, S.E. 1979. Temporal changes in the salinity and temperature requirements of tropical mussel larvae. *Proceedings of the World Mariculture Society*, 9, 549-566.
- Smale, H.J. and Buchan, P.R. 1981. Biology of *Octopus vulgaris* off the east coast of South Africa. *Marine Biology*. 65(1), 1-12.
- Smith, J. R., Peggy Fong, P., and Ambrose, R.F. 2009. Spatial patterns in recruitment and growth of the mussel *Mytilus californianus* (Conrad) in southern and northern California, USA, two regions with differing oceanographic conditions. *Journal of Sea Research*. 61, 165–173.
- Solomieu, V.B., Renault, T., and Travers, M. 2015. Mass mortality in bivalves and the intricate case of the Pacific oyster, *Crassostrea gigas*. *Journal of Invertebrate Pathology*. 131, 2-10.
- Sreedevi, P.R., Uthayakumar, V., Jayakumar, R., and Ramasubramanian, V. 2014. Influence of rearing water temperature on induced gonadal development and spawning behaviour of tropical green mussel, *Perna viridis*. *Asian Pacific Journal of Reproduction*. 3(3), 204-209.
- Strickland, J.D.H. and Parsons, T.R. 1972. *A practical handbook of seawater analysis*. Fisheries Research Board of Canada, Ottawa, pp. 11-26.
- Talling J.F. and Driver D. 1963. Some problems in estimation of Terrestrial algae: The British Physiological Society, Cambridge Taxonomic analysis of the genus *Anabaenopsis*. *Archiv fur Hydrobiologie*. 51, 3-24.
- Tamarin, A., Lewis, P., and Askey, J. 1974. Specialised cilia of the byssal attachment plaque forming region in *Mytilus californianus*. *Journal of morphology*. 142, 321-327.
- Tan, K.S. and Ransangan, J. 2014. A Review of Feeding behavior, Growth, Reproduction and Aquaculture Site Selection for Green-Lipped Mussel, *Perna viridis*. *Advances in Bioscience and Biotechnology*. 5, 462-469.
- Tuaycharden, S., Vakily, J.M., Saelow, A., and McCoy, E.W. 1988. Growth and maturation of the green mussel (*Perna viridis*) in Thailand. In McCoy, E.W., Chongpeepien, T. (Eds.), *Bivalve Mollusc Culture Research in Thailand*, pp. 88-101.
- Vakily, J.M., Tuaycharoen, S., and J. Nugranad. 1988. Analysis of length and weight characteristics of green mussel, *Perna viridis* from the Gulf of Thailand. *Asian Fisheries Science*. 1, 165-174.
- Vakily, J.M. 1989. The biology and culture of mussels of the Genus *Perna*. *International Centre for Living Aquatic Resources Management, Manila, Philippines*, pp. 1-63.
- Yap, C.K. 2012. *Mussel Watch in Malaysia Past, Present and Future*. Universiti Putra Malaysia Press, 2012.
- Zakaria, M. and Rajpar, M.N. 2015. Assessing the fauna diversity of Marudu Bay mangrove forest, Sabah, Malaysia, for future conservation. *Diversity*. 7, 137-148.
- Zardi, G.I., McQuaid, C.D., and Nicastro, K.R. 2007. Balancing survival and reproduction: seasonality of wave action, attachment strength and reproductive output in indigenous *Perna perna* and 9 invasive *Mytilus galloprovincialis* mussels. *Marine Ecology Progress Series*. 334, 155-163.
- Zbawicka, M., Sanko, T., Strand, J., and Wenne, R. 2014. New SNP markers reveal largely concordant clinal variation across the hybrid zone between *Mytilus* spp. in the Baltic Sea. *Aquatic Biology*. 21, 25-36.